open.michigan

Author(s): Michael Hortsch, Ph.D., 2009

License: Unless otherwise noted, this material is made available under the terms of the **Creative Commons Attribution – Noncommercial – Share Alike 3.0 License**: http://creativecommons.org/licenses/by-nc-sa/3.0/

We have reviewed this material in accordance with U.S. Copyright Law and have tried to maximize your ability to use, share, and adapt it. The citation key on the following slide provides information about how you may share and adapt this material.

Copyright holders of content included in this material should contact **open.michigan@umich.edu** with any questions, corrections, or clarification regarding the use of content.

For more information about **how to cite** these materials visit http://open.umich.edu/education/about/terms-of-use.

Any **medical information** in this material is intended to inform and educate and is **not a tool for self-diagnosis** or a replacement for medical evaluation, advice, diagnosis or treatment by a healthcare professional. Please speak to your physician if you have questions about your medical condition.

Viewer discretion is advised: Some medical content is graphic and may not be suitable for all viewers.





Citation Key

for more information see: http://open.umich.edu/wiki/CitationPolicy

Use + Share + Adapt

{ Content the copyright holder, author, or law permits you to use, share and adapt. }

@ PD-GOV	Public Domain – Government: Works that are produced by the U.S. Government. (17 USC §105)
Ø PD-EXP	Public Domain – Expired: Works that are no longer protected due to an expired copyright term.
@ PD-SELF	Public Domain – Self Dedicated: Works that a copyright holder has dedicated to the public domain.
(6) 2680	Creative Commons – Zero Waiver
(@) ov	Creative Commons – Attribution License
(C)) BY-SA	Creative Commons – Attribution Share Alike License
(C)) 8Y-NC	Creative Commons – Attribution Noncommercial License
(0) BY-NC-SA	Creative Commons – Attribution Noncommercial Share Alike License
S GNU-FDL	GNU – Free Documentation License

Make Your Own Assessment

{ Content Open.Michigan believes can be used, shared, and adapted because it is ineligible for copyright. }

Public Domain – Ineligible: Works that are ineligible for copyright protection in the U.S. (17 USC § 102(b)) *laws in your jurisdiction may differ

{ Content Open.Michigan has used under a Fair Use determination. }

Fair Use: Use of works that is determined to be Fair consistent with the U.S. Copyright Act. (17 USC § 107) *laws in your jurisdiction may differ

Our determination **DOES NOT** mean that all uses of this 3rd-party content are Fair Uses and we **DO NOT** guarantee that your use of the content is Fair.

To use this content you should do your own independent analysis to determine whether or not your use will be Fair.

Histology of the Peripheral Nervous System

Michael Hortsch, Ph.D. Department of Cell and Developmental Biology University of Michigan

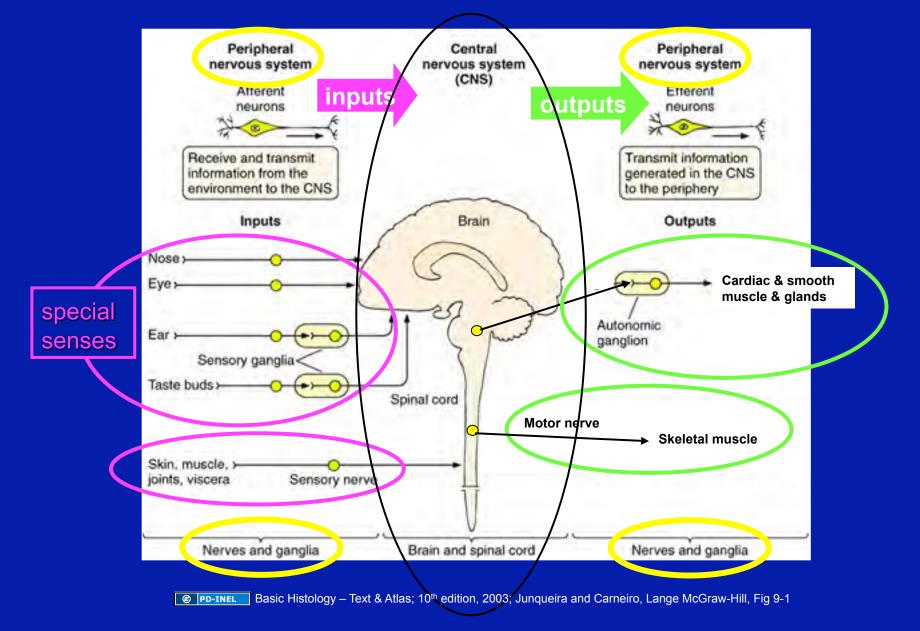


Winter, 2009

Objectives of PNS Histology:

- Discuss the general division/differences between CNS and PNS
- Appreciate the subdivision into somatic and autonomic nervous system
- Learn about the cellular components and the structural attributes of neuronal cells
- Discuss synaptic connections, using the motor end plate as an example
- Study the formation of the axonal myelin ensheathment
- Compare the histological features of myelinated and unmyelinated axons/nerves
- Recognize nerves in histological sections
- Identify the different connective tissue layers that are associated with nerves
- Understand the different organizational plans that are adopted by neuronal cells
- Identify and compare autonomic and sensory ganglia
- Learn about the basic histological features of the spinal cord
- Understand the organization and functions of mechanosensory receptors and neuromuscular spindles and be able to recognize them

Structural Organization of the Nervous System



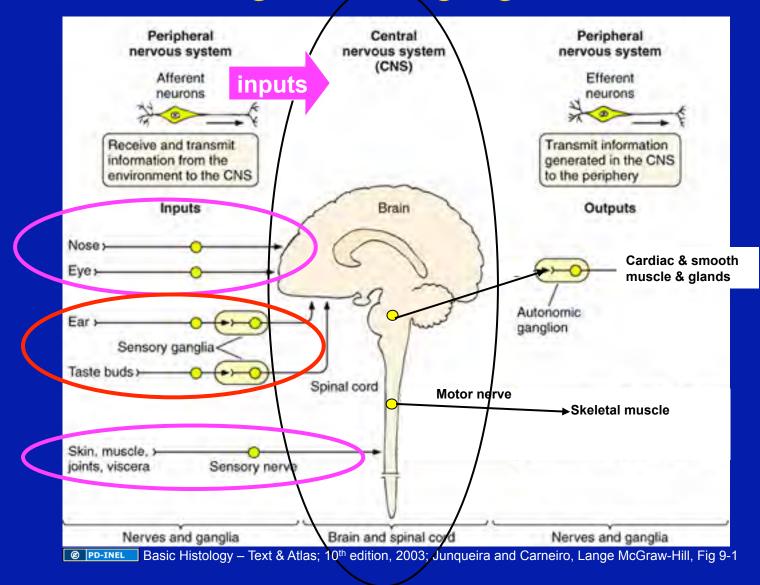
Functional Organization of the Nervous System

- 1. Somatic (conscious afferent* and efferent, voluntary motor control)
- 2. Autonomic (unconscious efferent, involuntary motor control of internal organs to maintain homeostasis)
 - a. Sympathetic thoracolumbar division
 - b. Parasympathetic craniosacral division

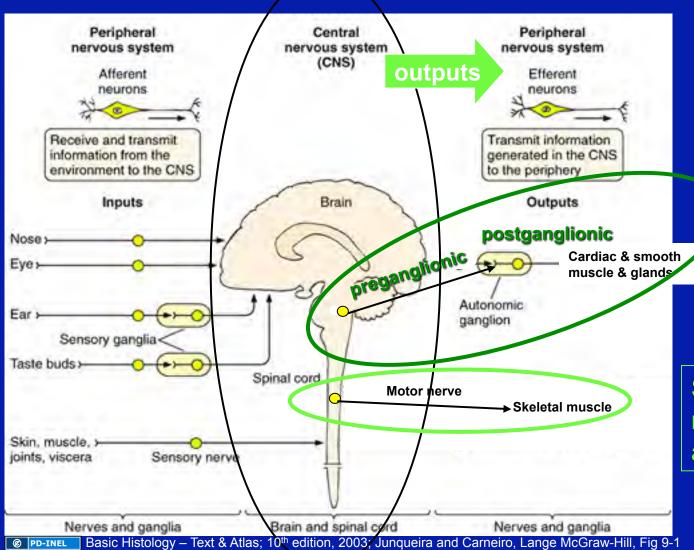
* Somatic afferents = sensory fibers from skin, muscle, joints, tendons.

Visceral afferents = sensory fibers from visceral organs; some result in conscious sensations, but others do not. However, they are not considered part of the autonomic nervous system, which is entirely efferent.

Perikarya of sensory neurons are in the PNS, often organized in ganglia



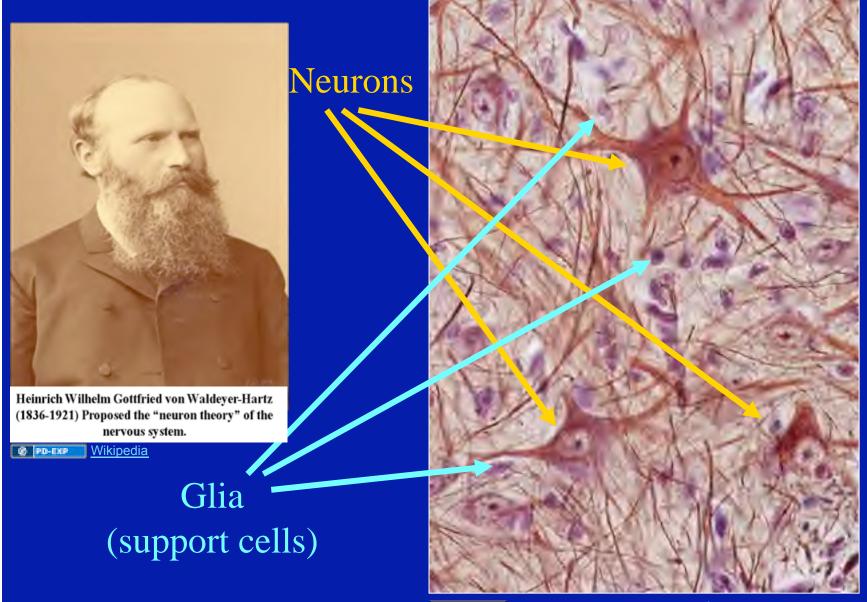
Motor neuron perikarya: somatic vs. autonomic



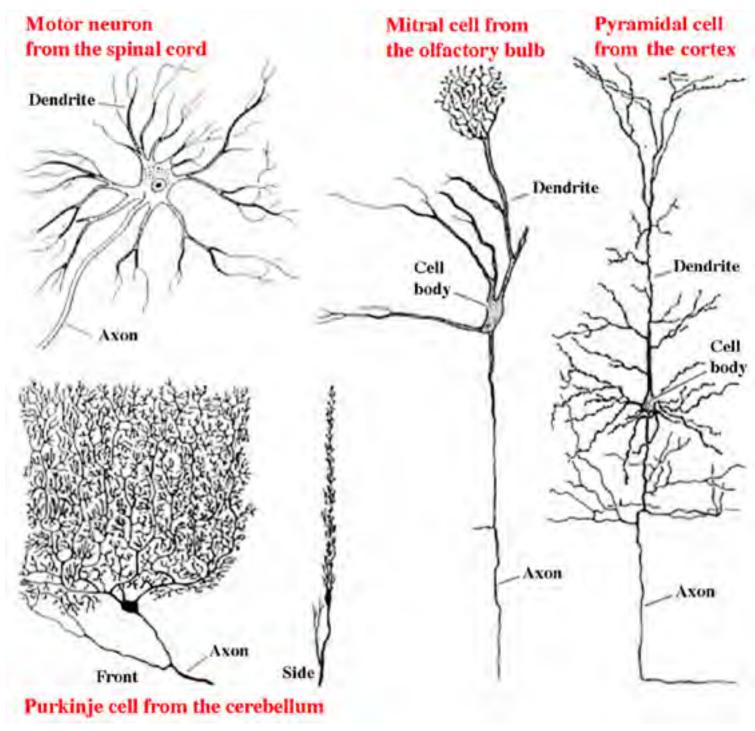
Autonomic efferents = two neuron chain, with 1st neuron in the CNS and 2nd neuron in PNS ganglia

Somatic motor neuron perikarya are in the CNS

Cellular Components of the Nervous System



© PD-INEL Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.4d

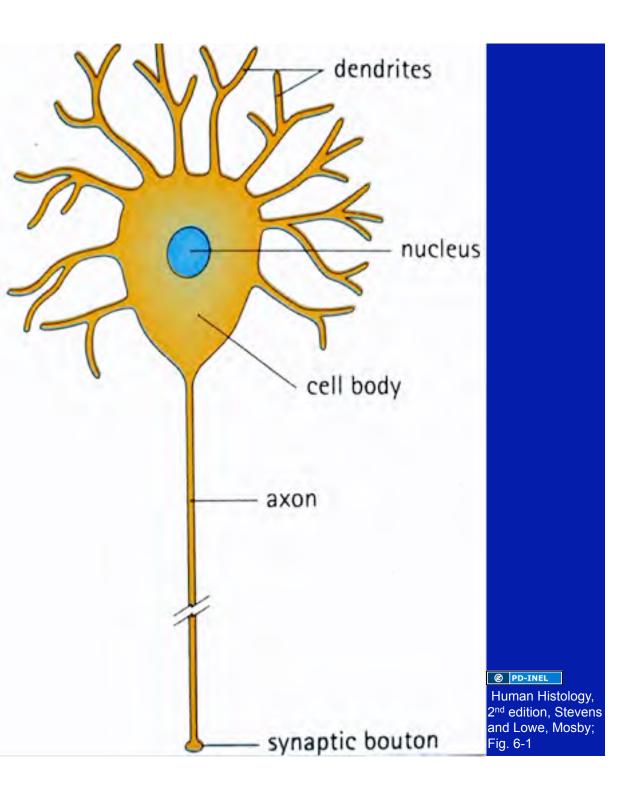


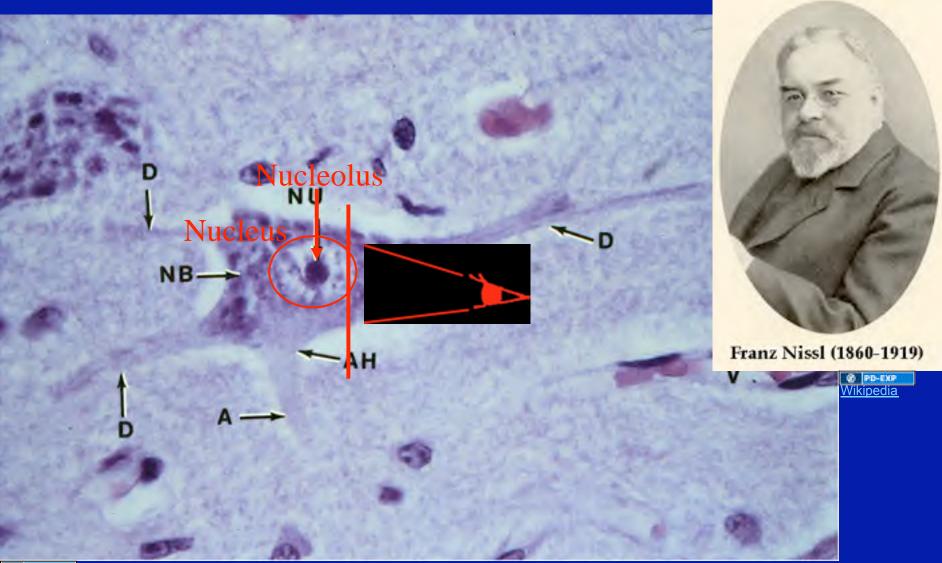
Neurons come in many shapes and forms

PD-INEL Neuron to Brain, 3rd edition, 1992; Nicholls, Martin and Wallace, Sinauer; Fig 6

Generic neuron

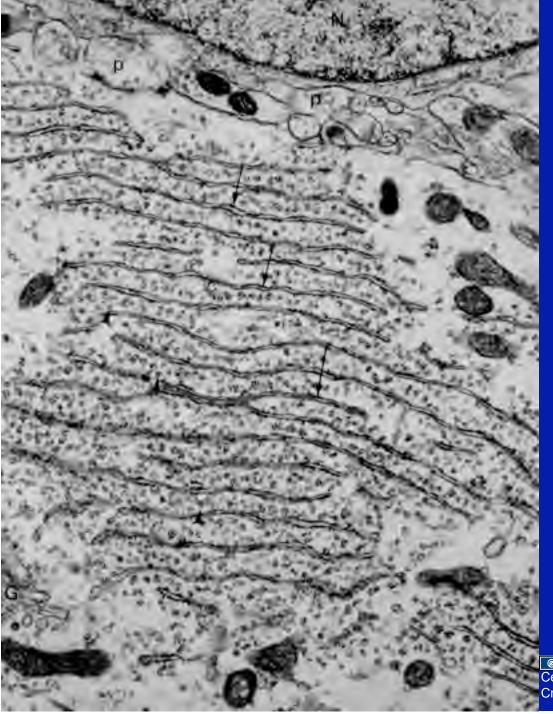
The cell body of a neuron is referred to as the <u>soma</u> or <u>perikaryon</u>





© PD-INEL Color Atlas of Basic Histology; 1993; Berman; Appelton and Lange; Fig 6-4

Motor neuron with Nissl substance

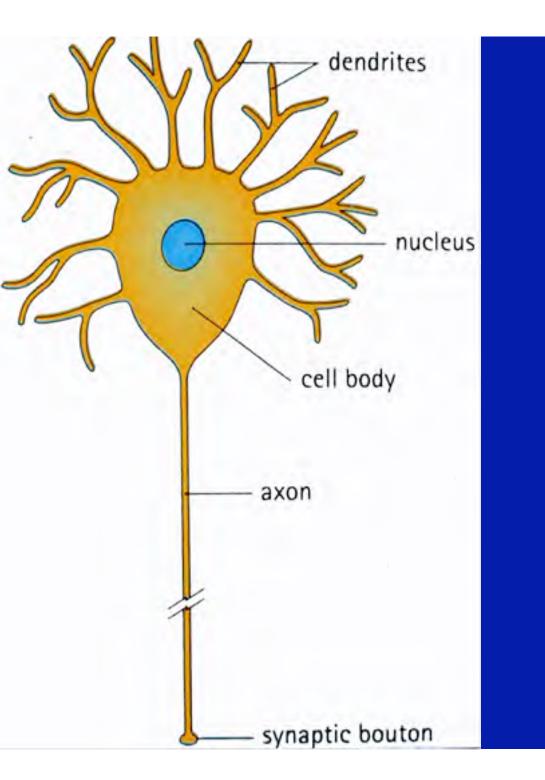


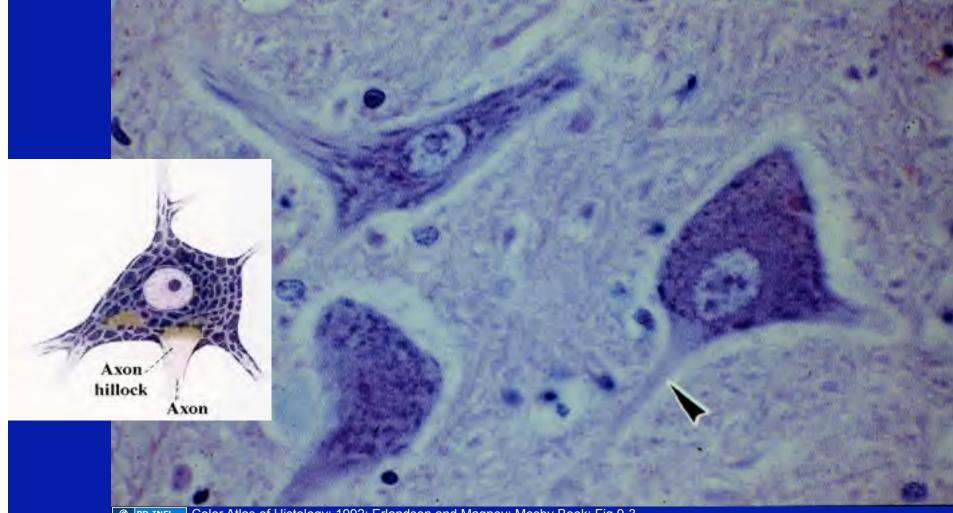
Nissl substance is rough endoplasmic reticulum

© PD-INEL Cell and Tissue Ultrastructure – A Functional Perspective; 1993; Cross and Mercer, Freeman and Co.; Page 127 Neurons have dendritic and <u>axonal</u> extension

The Law of Dynamic <u>Polarization</u> states that neuronal signals only travel in one direction, from dendrites to the axon. In humans axons can be up to 1.5 meters in length. In a whale axonal length can reach up to 40 meters.

> Human Histology, 2nd edition, Stevens and Lowe, Mosby; Fig. 6-1 @ PD-INEL

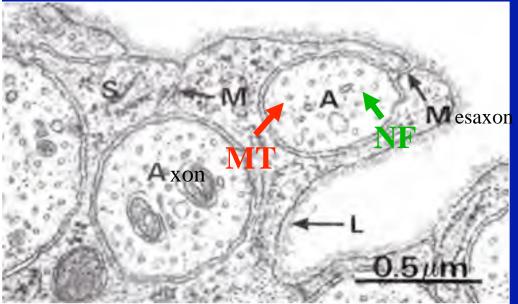




© PD-INEL Color Atlas of Histology; 1992; Erlandsen and Magney; Mosby Book; Fig 9-3

Nissl substance is found in the neuronal cell body and dendrites, but not in the axon and the axon hillock or axon initial segment. The ability of neurons to synthesize proteins at growth cones and at the presynaptic terminus is <u>very</u> limited.

How do axons grow to reach their targets and how is an adult axon maintained and supplied?

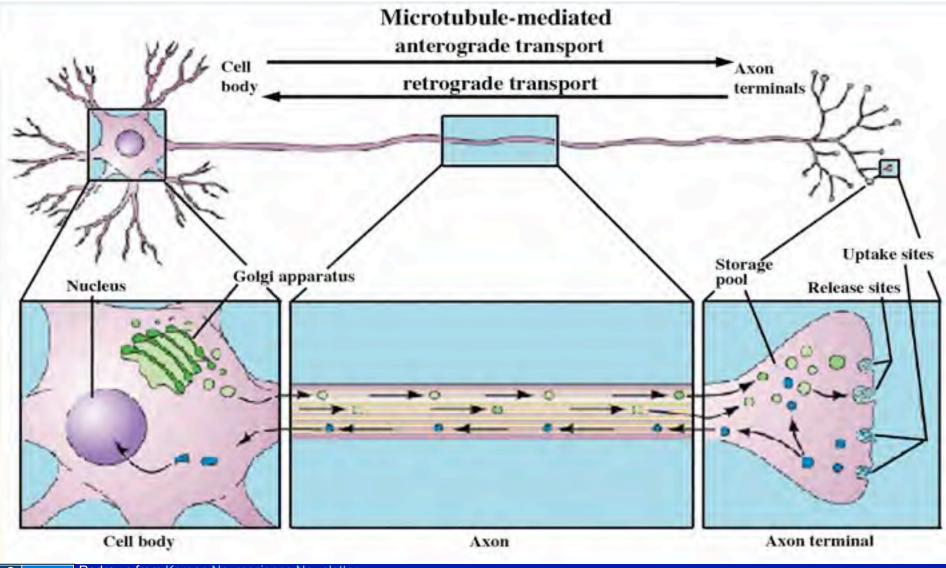


Axonal cytoskeletal elements play a central role in the two types of transport along axons. As shown in this electron micrograph, axons contain two classes of cytoskeletal elements, neurofilaments (NF), which are intermediate filaments, and microtubules (MT).

PD-INEL Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.5c

<u>Slow axonal transport</u> (0.2-8 mm/day) transports many proteins along the axons. The mechanism of slow axonal transport is still controversial (current Stop-Go model).

<u>Fast axonal transport</u> (up to 400 mm/day) is the main mechanism to move cell organelles and membrane vesicles along the axon.



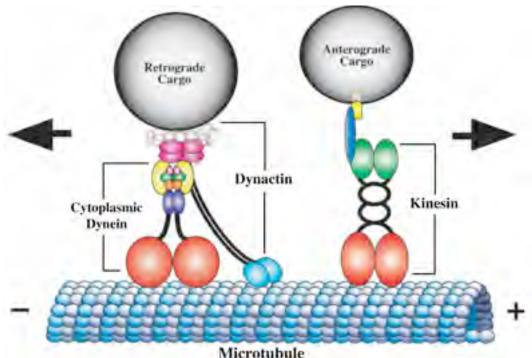
PD-INEL Redrawn from Korean Neuroscience Newsletter

Fast axonal transport goes in both directions (anterograde and retrograde) and relies on the axonal microtubule cytoskeleton.



PD-INEL J.E. Lochner, M. Kingma, S. Kuhn, C.D. Meliza, B. Cutler, B.A. Scalettar

Microtubule-mediated transport of secretory granules along the axon of a neuron. The majority of granules move toward the growth cone (anterograde transport), but some move away from the growth cone (retrograde transport). There are also some granules that reverse direction, move intermittently, or stall. In this neurite, there is a partial build up of granules in the growth cone; this build up would have become more extensive with time due to the net anterograde granule flux.



Anterograde microtubulemediated transport is mediated by kinesins and retrograde transport by dyneins. The microtubule-mediated transport enables the tip of an axon to grow during development and to regeneration. During this time is referred to as a growth cone.



Snail growth cone stained for actin and microtubules.

© PD-INEL Reproduced from The Journal of Cell Biology, 2002, 157 (5) by copyright permission of The Rockefeller University

Drosophila growth cone

Ø PD-INEL David Van Vactor, Havard University

Originally from Duncan JE, Goldstein LSB Ø PD-INEL (2006) The Genetics of Axonal Transport and Axonal Transport Disorders. PLoS Genet 2(9): Pages 1275ce -84.

Once a growth cone reaches its target, it might form synapses with its target cell. Synapses are predominantly supplied and maintained by fast, microtubule-mediated transport.

Press

Synapses can form between many different parts of neurons and between a neuron and a non-neuronal cell, e.g., a muscle or a secretory cell. axo-dendritic axo-somatic axo-axonic axo-axonic

PD-INEL Human Histology, 2nd edition, Stevens and Lowe, Mosby; Fig 6.7

A single neuron can receive activating or inhibiting inputs from thousands of synaptic connections.

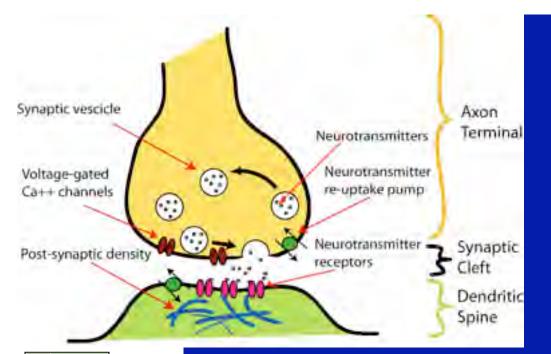
Ø PD-INEL

Panel B courtesy of Olaf Mundigl and Pietro de Camilli in The Molecular Biology of the Cell by B. Alberts et al., 4th edition, 2002, Garland Science

Images of synapses and motor neuron cell body in spinal cord removed

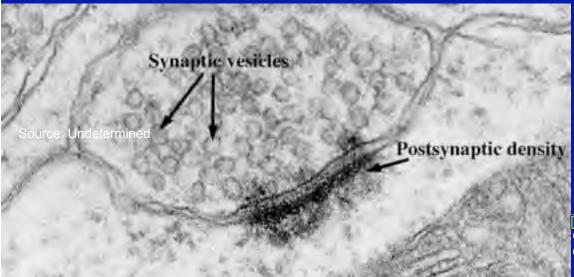
Source of Removed Image: The Molecular Biology of the Cell by B. Alberts et al., 4th edition, 2002, Garland Science Fig. 11-38 A

Motor neuron cell body in the spinal cord



③ GNU-FDL

<u>Wikipedia</u>



ORIGINAL TOP IMAGE Diagram of synapse downloaded from http://fantastrid.googlepages.com/anatomydrawings_by Astrid Vincent Andersen Web page http://fantastrid.googlepages.com/homedk

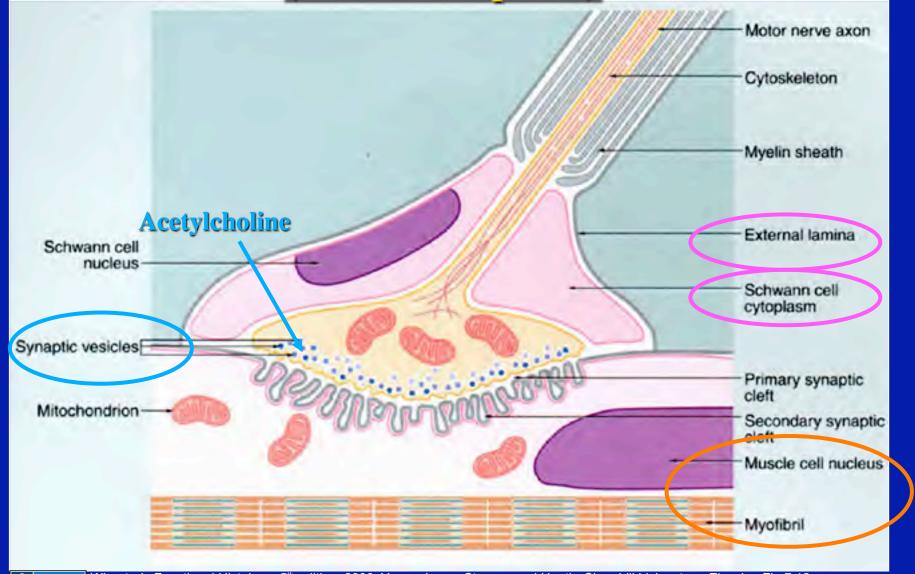
At a chemical synapse neurotransmitter release is triggered by the influx of Ca²⁺ and postsynaptic neurotransmitter receptors receive the signal.

Ø PD-INEL

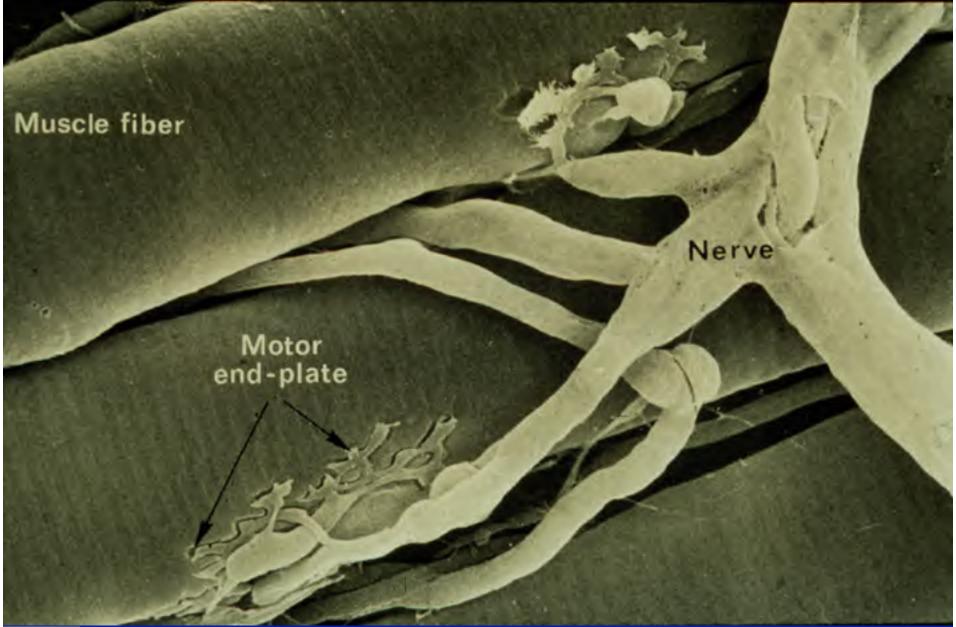
Cell and Tissue Ultrastructure – A Functional Perspective by Cross and Mercer; 1993; Freeman and Co. Page 135

Neuromuscular junction

(motor endplate)



© PD-INEL Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.12a



© PD-INEL Color Atlas of Histology; 1992; Erlandsen and Magney; Mosby Book; Fig 8-18

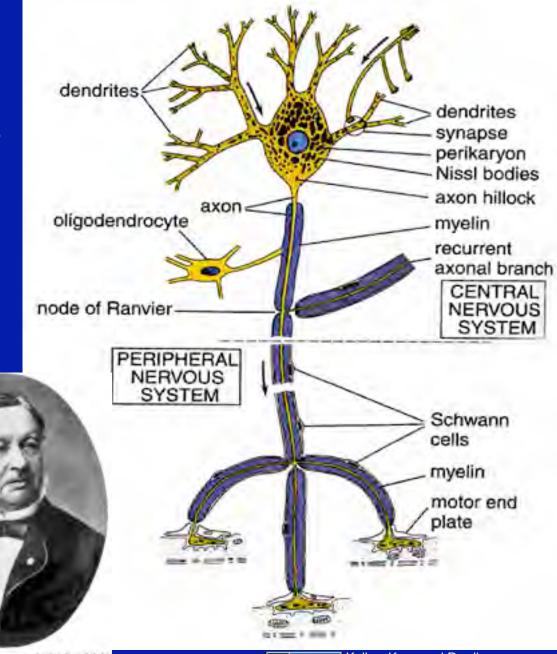
Scanning EM of motor endplate on a muscle fiber

Motor endplates on skeletal muscle fibers



© PD-INEL Photograph by AK Christensen from slide by Ray Truex, Dept of Anatomy, Temple Univ. School of Medicine

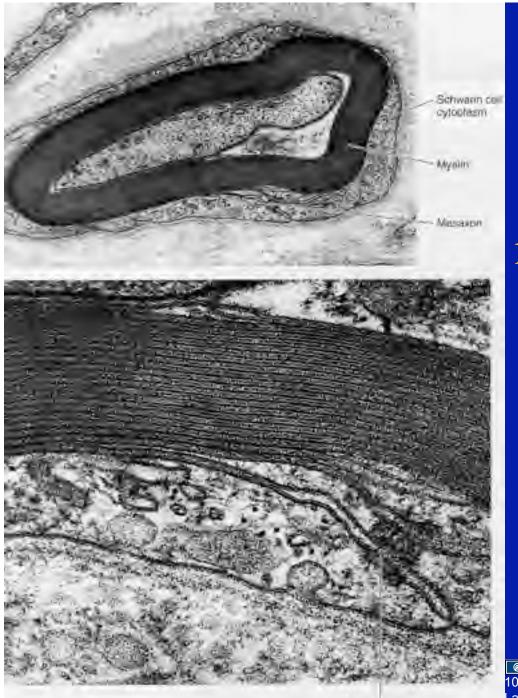
Myelination in the CNS involves oligodendrocytes and Schwann cells in the PNS



Theodor Schwann (1810-1882)

PD-INEL Wikipedia

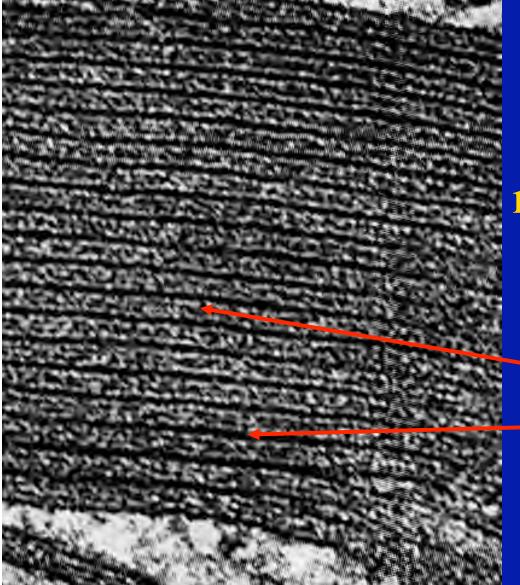
PD-INEL Kelley, Kaye and Pawlina, "Histology, a Text and Atlas," 4th ed., page 284. Neuron-Ross4-284.tif.



Outor mesawori

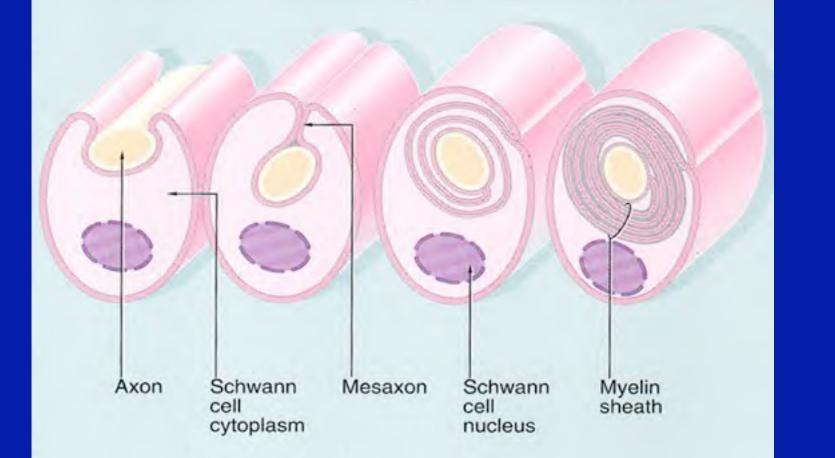
This electron micrograph of a single myelinated axon shows a series of lighter (intraperiod) and darker (major dense) lines

PD-INEL Basic Histology – Text & Atlas; 10th edition, 2003; Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-30



This electron micrograph of a single myelinated axon shows a series of lighter (intraperiod) - and darker (major dense) lines

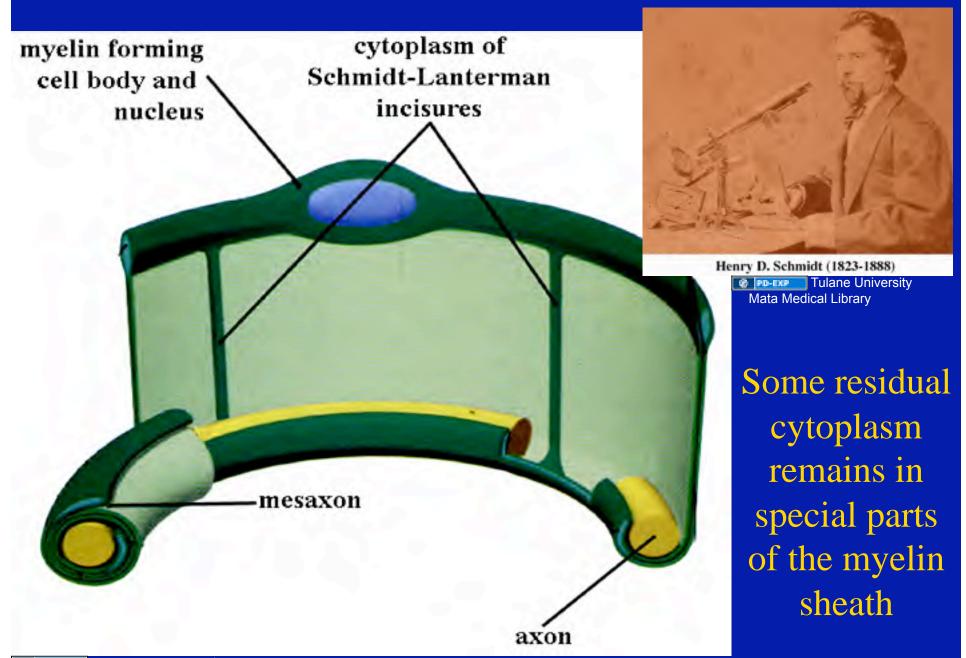
PD-INEL Basic Histology – Text & Atlas; 10th edition, 2003; Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-30



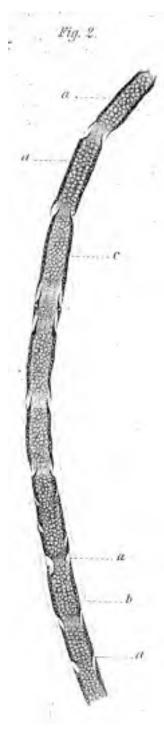
PD-INEL Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.6a

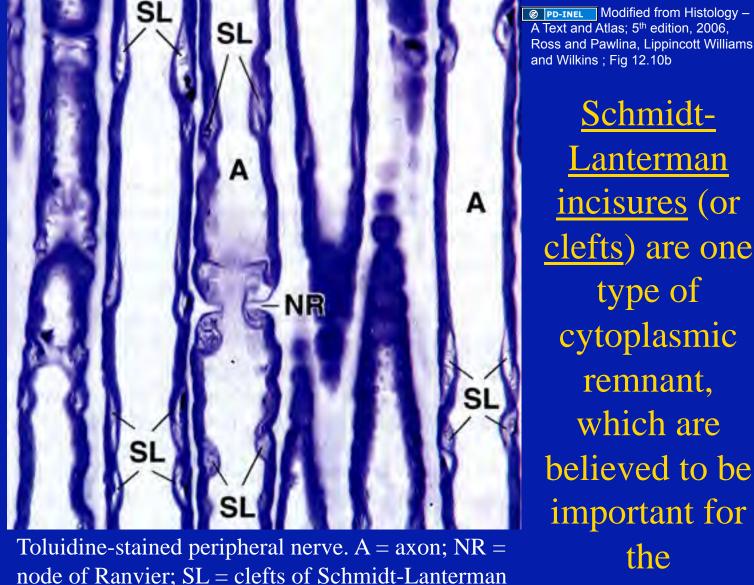
Myelination is a dynamic process, which involves the ensheathment of the the axon by the glial cell and subsequently the extrusion of cytoplasm from parts of the glial cell. Adhesive proteins on the cytoplasmic and the extracellular side of the plasma membrane contribute to a tight apposition of the lipid bilayers. ^{Original Image:} Histology-A Text and Atlas by M.H. Ross

Histology-A Text and Atlas by M.H. Ross and W. Pawlina; 5th edition, 2006, Lippincott Williams and Wilkins, Fig 12.11



© PD-INEL Human Histology, 2nd edition, Stevens and Lowe, Mosby; Fig. 6.9





type of cytoplasmic remnant, which are believed to be

node of Ranvier; SL = clefts of Schmidt-Lanterman

important for the maintenance of the myelin sheet.

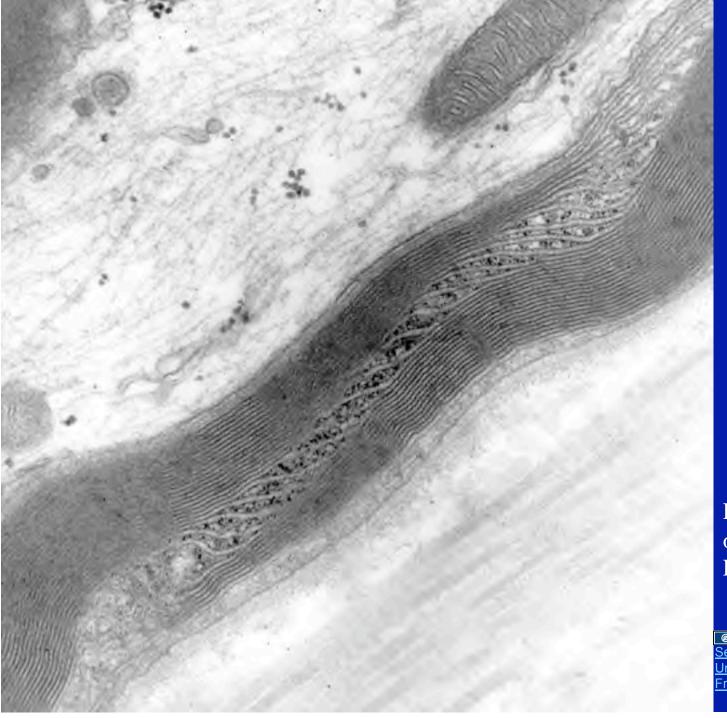
Modified from Histology –

Schmidt-

Lanterman

incisures (or

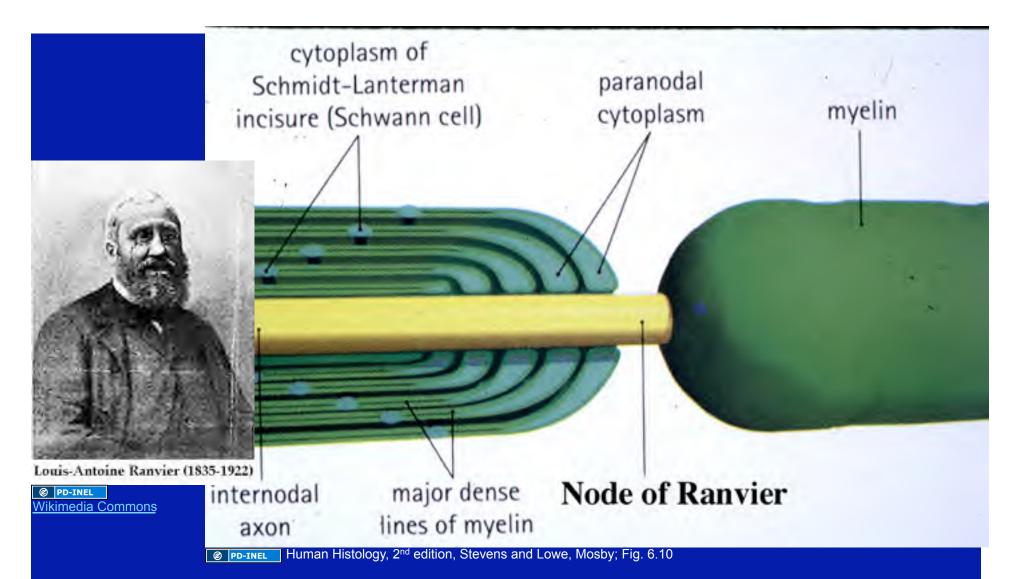
clefts) are one



Schmidt-Lanterman incisures (or clefts) are one type of cytoplasmic remnant, which are believed to be important for the maintenance of the myelin sheet.

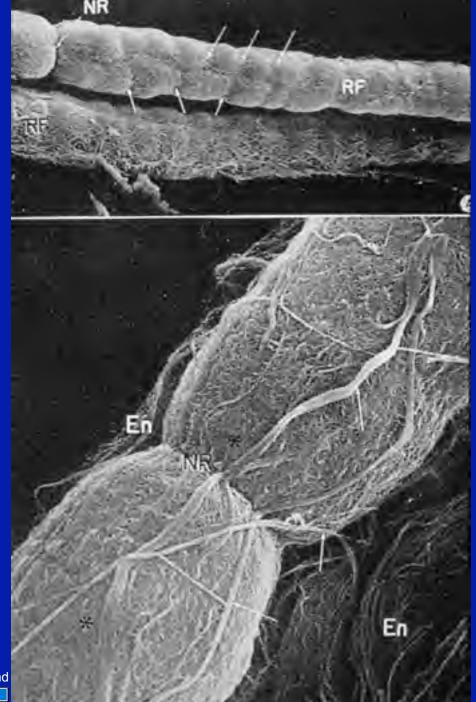
Electron micrograph of a Schmidt-Lanterman incisure

© PD-INEL Service De Neurologie, Hôpital Universitaire Dupuytren, Limoges, France



<u>Nodes of Ranvier</u> are areas of the myelinated axon that are not covered by the myelin sheath. They are bordered by paranodal regions, which form <u>paranodal junctions</u> with the axonal plasma membrane and also retain some Schwann cell cytoplasm.

Scanning EMs depicting nodes of Ranvier



Color Atlas of Histology; 1992; Erlandsen and Magney; Mosby Book; Fig 9-15 @ PD-INEL

Paranodal junctions

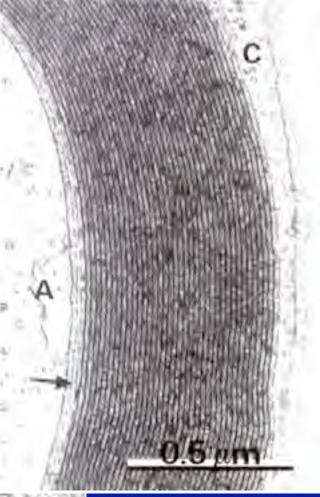
Transmission EM with node of Ranvier and paranodal region

© PD-INEL Color Atlas of Histology; 1992; Erlandsen and Magney; Mosby Book; Fig 9.14

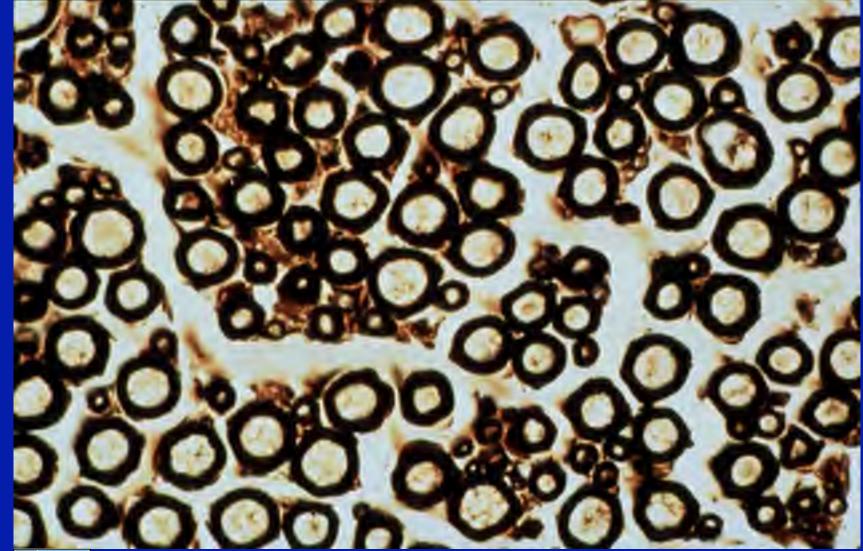
Myelinated Nerve Fiber

The increased lipid content of the myelin sheath provides electrical insulation for the underlying axon.



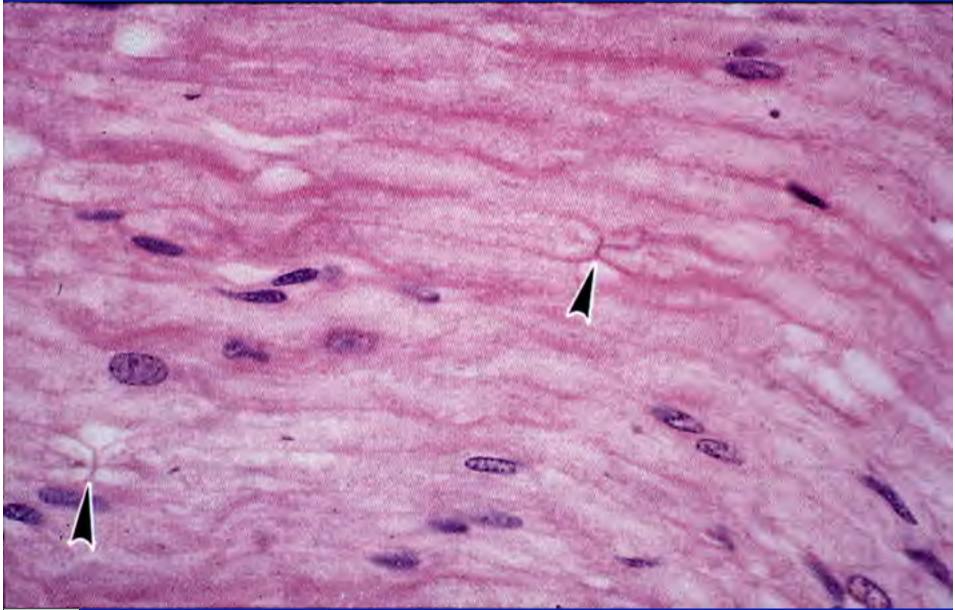


PD-INEL Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig. 7.6 A, axon; S, Schwann cell nucleus.



PD-INEL Japanese Kodachrome slide set, Slide 1084

Silver-stained cross section of a myelinated nerve



© PD-INEL Color Atlas of Histology; 1992; Erlandsen and Magney; Mosby Book; Fig 9-13

Nodes of Ranvier in a longitudinal nerve section

Each Schwann cell myelinates a single internode

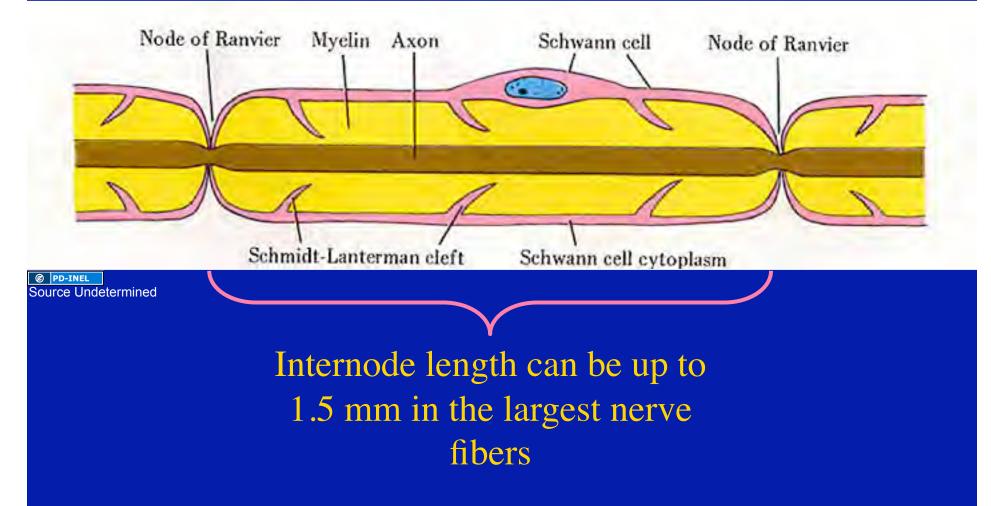


Diagram of myelinated axon and action potential propagation removed

Original Source Removed Modified from <u>Neuroscience by D. Purves et al.,</u> 2001, 2nd ed., Sinauer Fig. 3.13

Ion channels are concentrated at the nodes of Ranvier and the myelin sheath acts as an electrical insulator. This allows for <u>saltatory</u> conductance of the action potential and increases the transmission speed of the nerve impuls.

Depending on the diameter of the axon, myelination increases the action potential speed approximately 5 to 50fold (up to >110 m/sec).



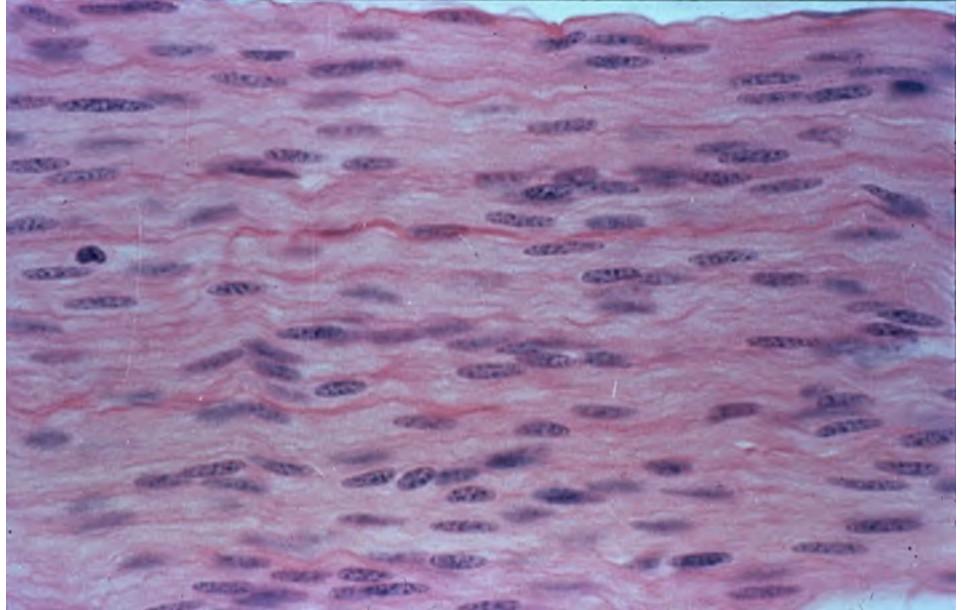
PD-INEL Human Histology, 2nd edition, Stevens and Lowe, Mosby; Fig. 6-21

One Schwann cell can ensheath multiple axons, but myelinates only one axon

Small diameter nerve fibers are non-myelinated

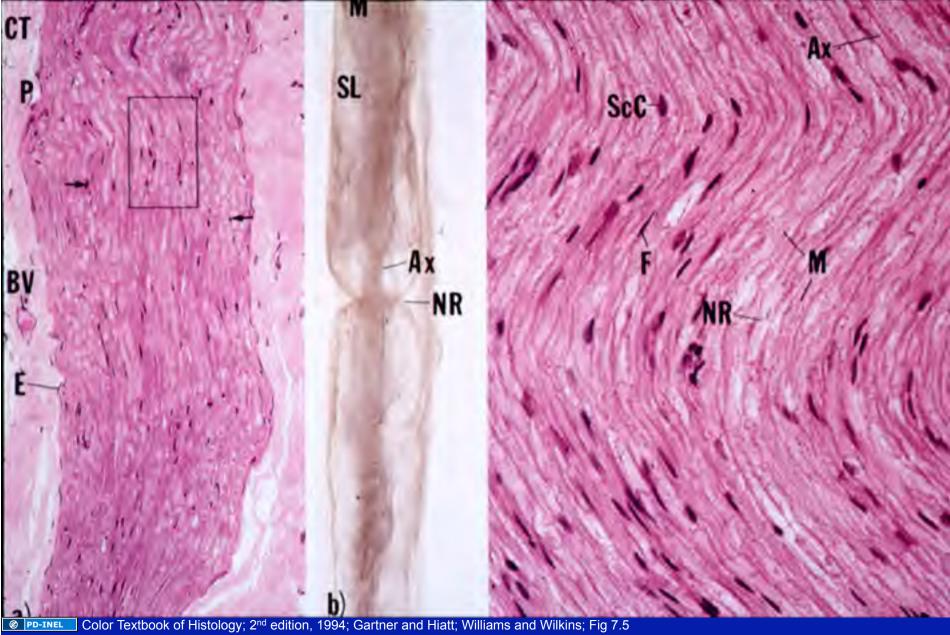


PD-INEL Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.5b



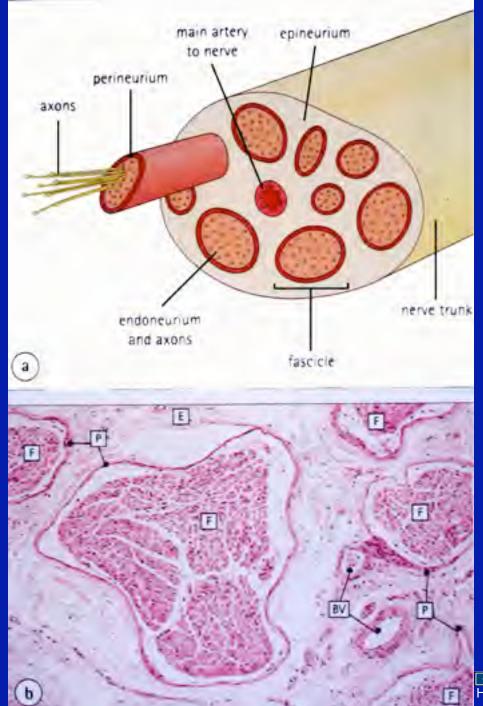
© PD-INEL Japanese slide set, Humio Mizoguti, Department of Anatomy, Kobe University School of Medicine, Slide #1091

Longitudinal section of an unmyelinated nerve



Ø PD-INEL

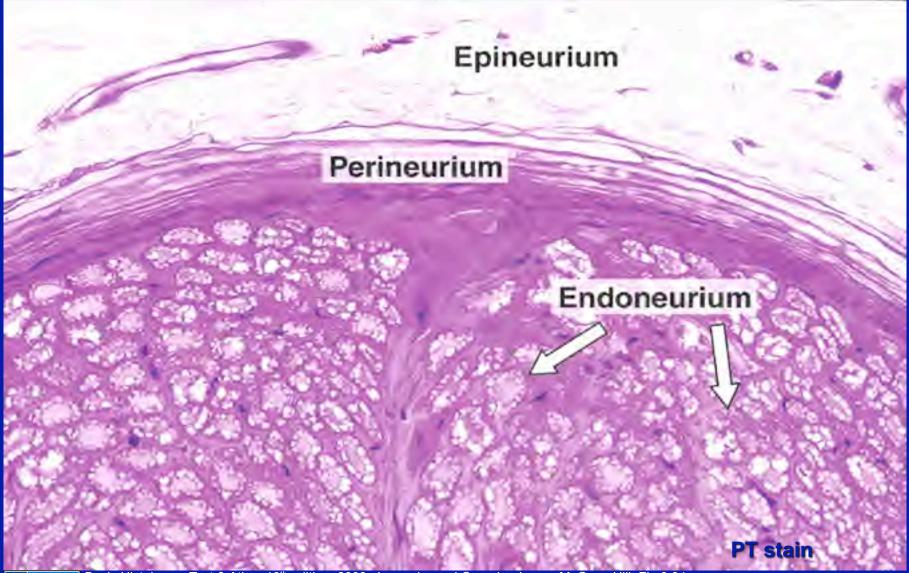
Wavy appearance of nerves



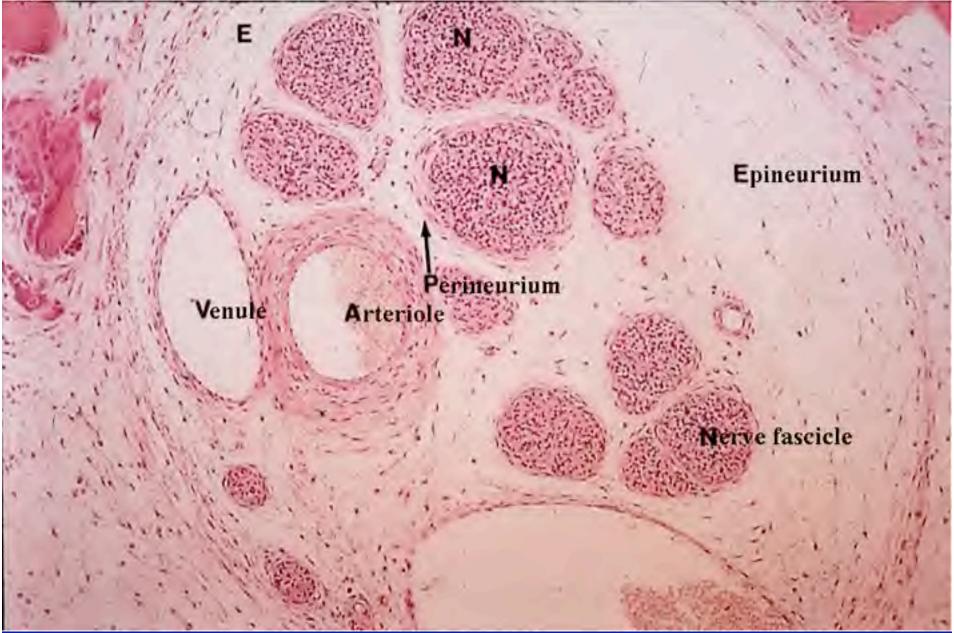
Connective tissue layers found in nerves: endoneurium surrounds axons, perineurium axon fascicles and epineurium the entire nerve

WITHEL HUMAN HISTOLOGY, 2nd edition, Stevens and Lowe, Mosby; Fig. 6.20

Connective tissue layers in a peripheral nerve. Tight junctions between perineurium cells form a important isolating barrier.

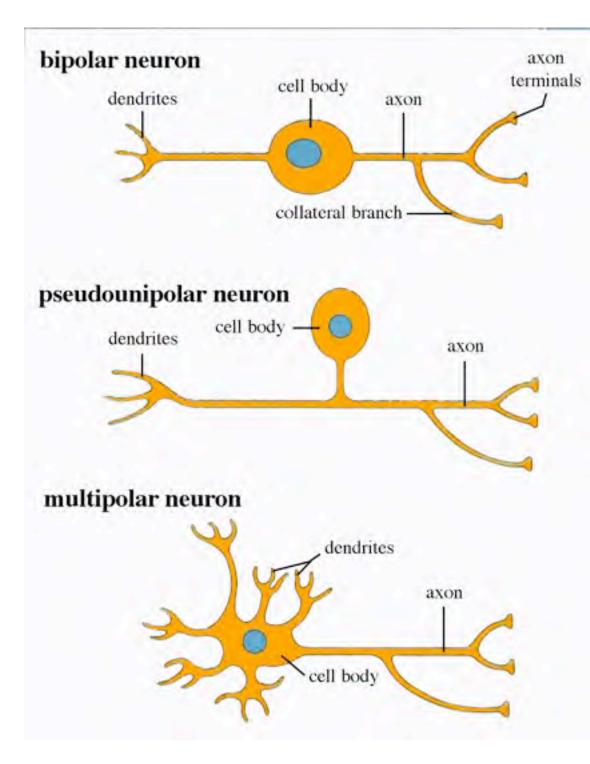


© PD-INEL Basic Histology – Text & Atlas; 10th edition, 2003; Junqueira and Carneiro, Lange McGraw-Hill, Fig 9.34



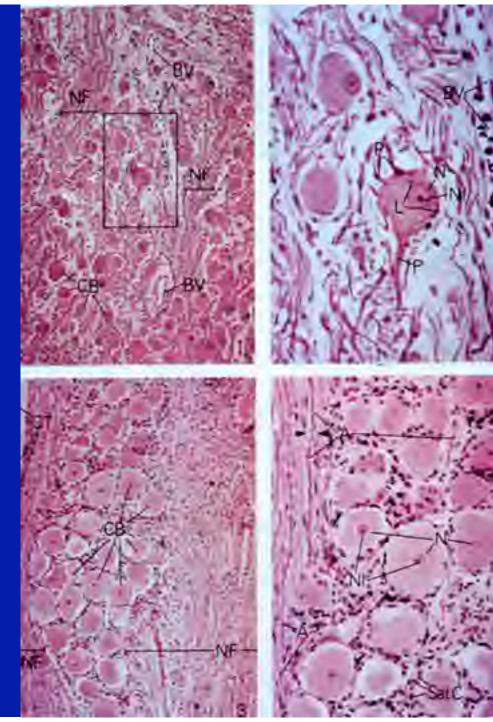
© PD-INEL Color Atlas of Basic Histology; 1993; Berman; Appelton and Lange; Fig 6.15

Connective tissue layers in a peripheral nerve cross section



Three different basic types of neuronal structure

PD-INEL Human Histology, 2nd edition, Stevens and Lowe, Mosby; Fig. 6.3



Autonomic ganglia with multipolar neurons are less organized than sensory ganglia (dorsal root ganglia) with pseudounipolar neurons.

PD-INEL
Histology – A Text and Atlas; 5th edition, 2006, Ross and Pawlina, Lippincott Williams and Wilkins; Plate 23

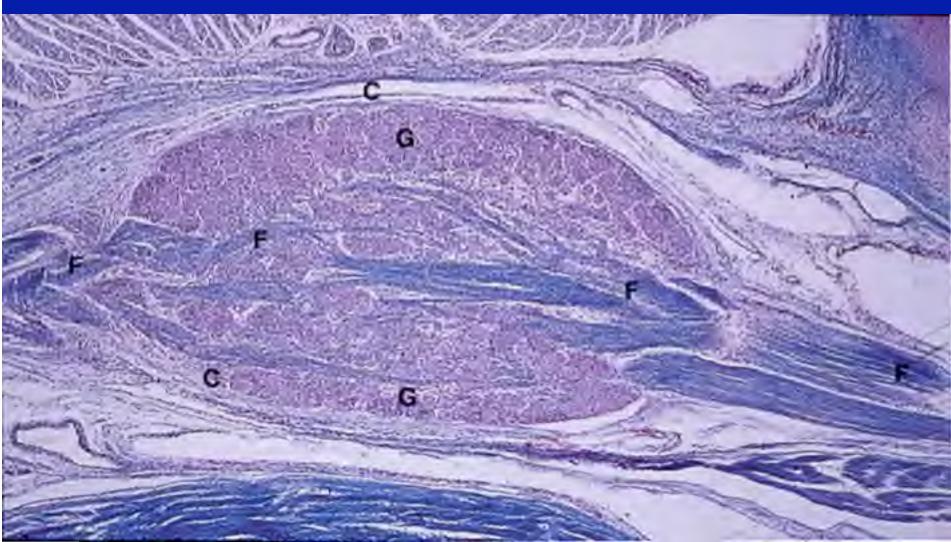
Sensory Ganglia

- Two types: spinal (dorsal root) and cranial ganglia associated with spinal and cranial nerves, respectively
- Contain large sensory neurons and abundant small glial cells, called satellite cells
- Sensory neurons are pseudounipolar



PD-INEL Source Undetermined

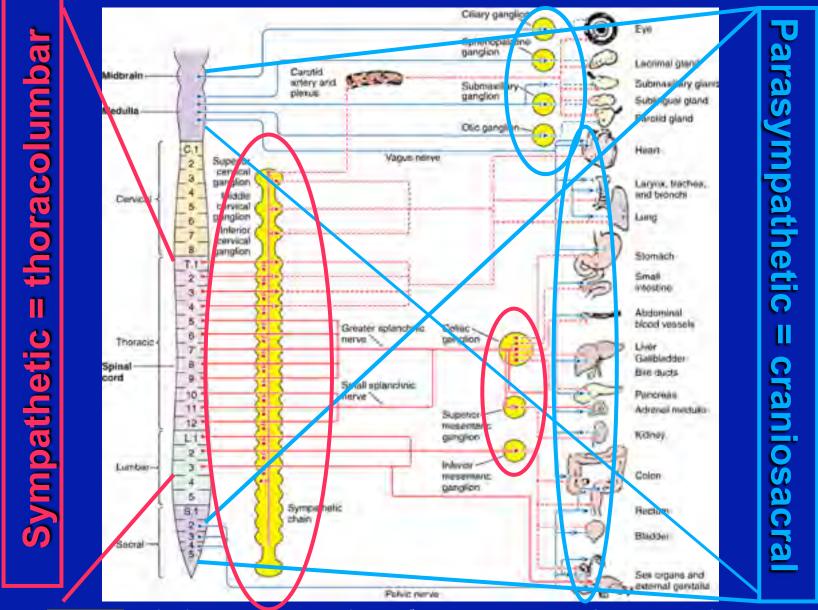
Dorsal root ganglion with pseudounipolar neurons



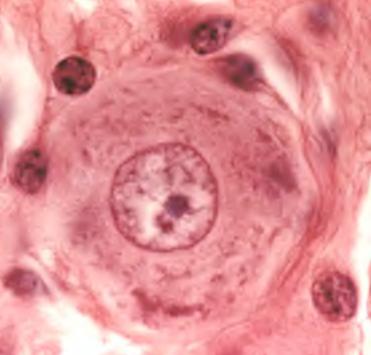
© PD-INEL Source Color Atlas of Basic Histology; 1993; Berman; Appelton and Lange; Fig 6-10

Luxol blue staining of dorsal root ganglion

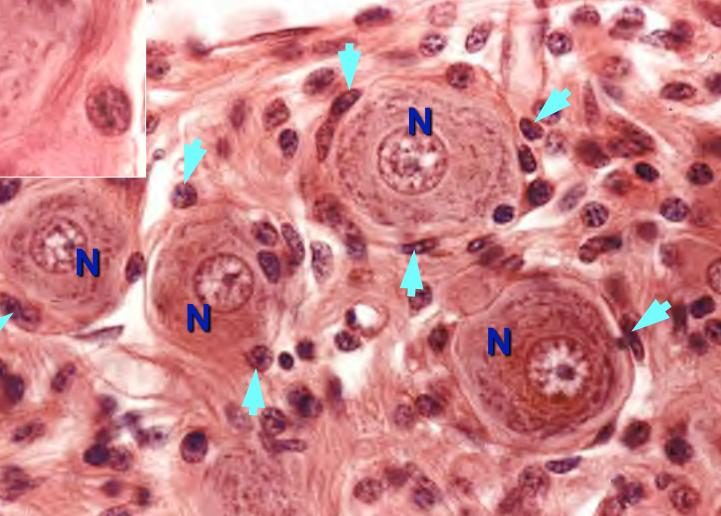
Efferent autonomic pathways



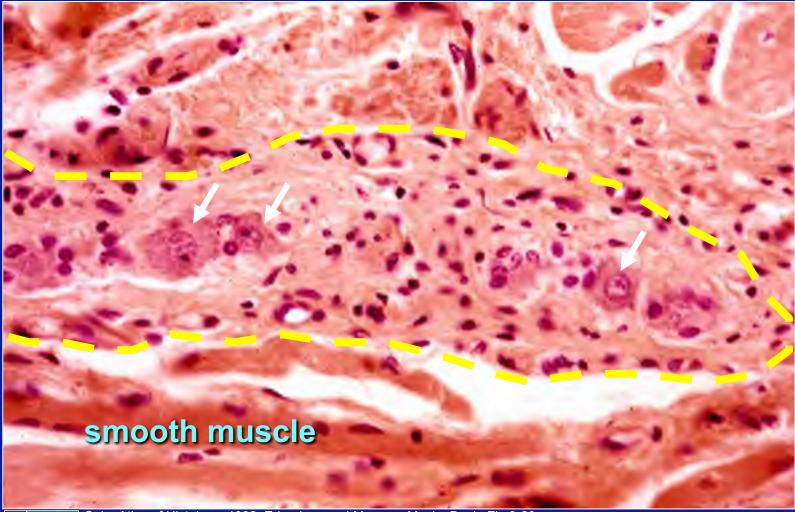
Sector 2013 (a) 10 (a) 10 (c) 10 (c)



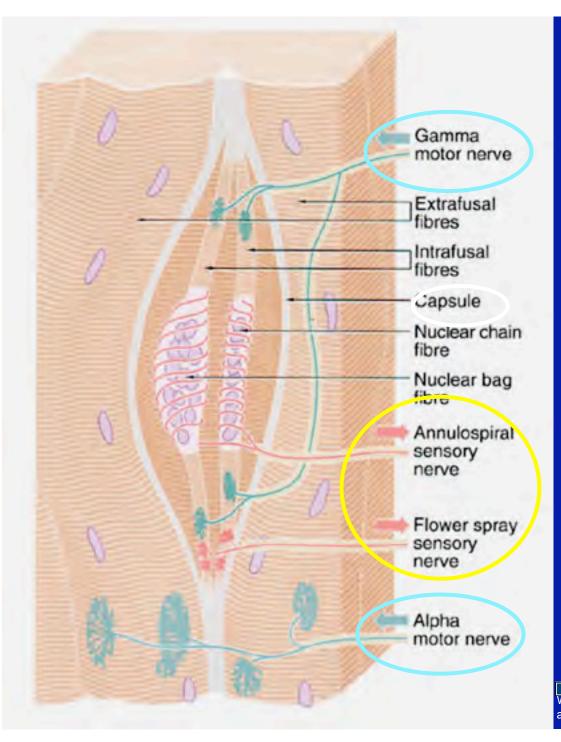
 PD-INEL Japanese slide set, Humio Mizoguti, Department of Anatomy, Kobe University School of Medicine, Slides #1069a and #1069b Autonomic Neurons in Sympathetic Ganglia are multipolar. These neurons are surrounded by satellite cells (glia cells marked by blue arrow heads).



Parasympathetic ganglia are located within or near their effector organs



© PD-INEL Color Atlas of Histology; 1992; Erlandsen and Magney; Mosby Book; Fig 9-20



Neuromuscular spindle (of Kühne)

Gamma motor nerve fibers innervate the intrafusal fibers



Wilhelm Friedrich Kühne (1837-1900) PD-EXP Wikipedia

Two-neuron stretch reflex

Sensory fiber endings are located on modified, small (intrafusal) muscle fibers

Alpha motor nerve fibers

Ø PD-INEL

Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.33a

Neuromuscular spindle

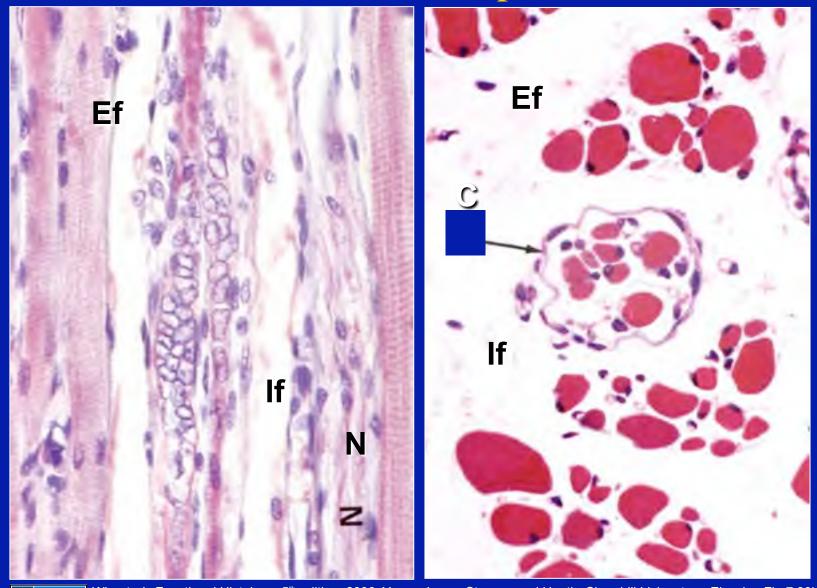
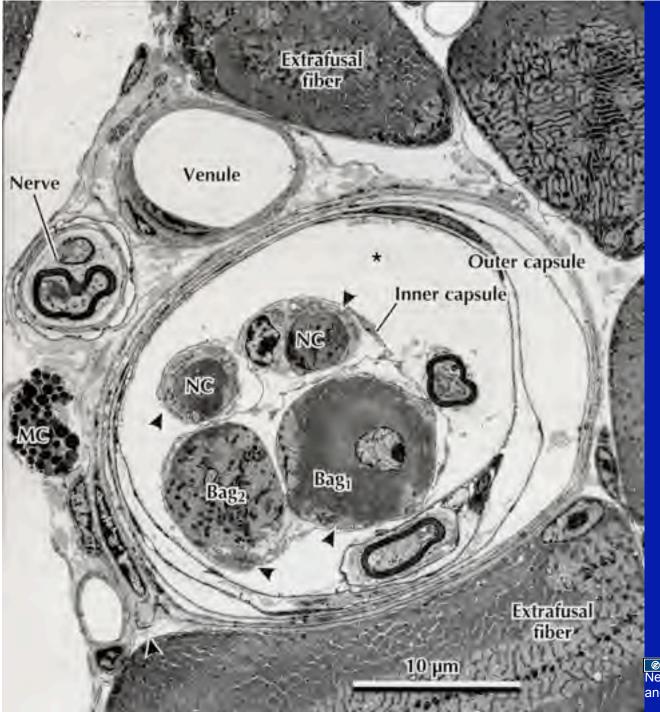


 Image: PD-INEL
 Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.30b

 Longitudinal section
 Transverse section



EM of a muscular spindle in the equatorial region

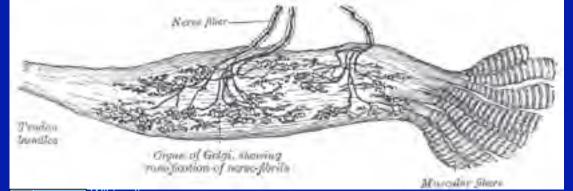
® PD-INEL Netter's Essential Histology; 2008; Ovalle and Nahirney; Elsevier; Page 467



Sensory mechanoreceptor at the tendon-muscle junction:

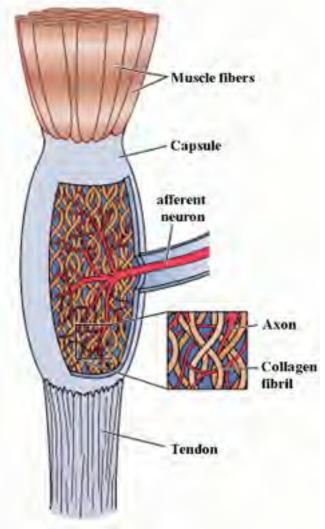
<u>Organ of Golgi or</u> <u>neurotendinous spindle</u>

Camillo Golgi (1843-1926) Camillo Golgi (1843-1926) Modified from Wikimedia Commons

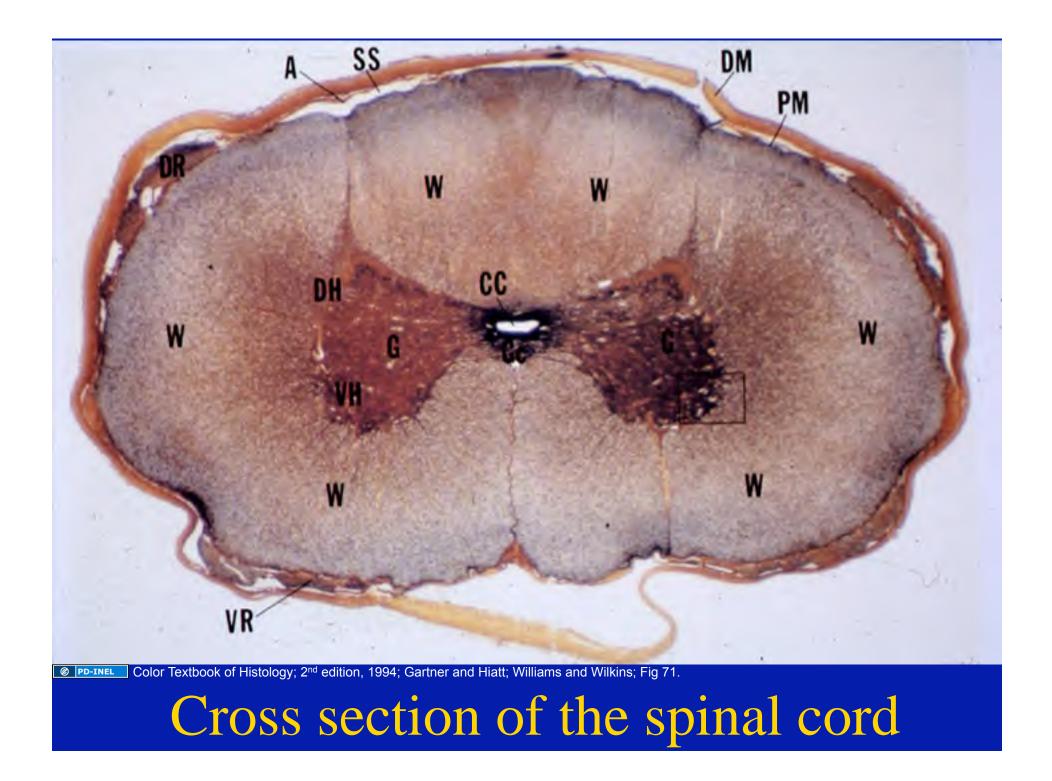


© PD-INEL Wikipedia

This organ of Golgi is an encapsulated stretch receptor. The capsule contains collagen fibers and endings of a single nerve fiber that is connected with interneurons in the spinal cord. Stretching forces will result in a depolarization of the axon and an inhibitory muscle reflex to protect muscles and tendons from excessive force.

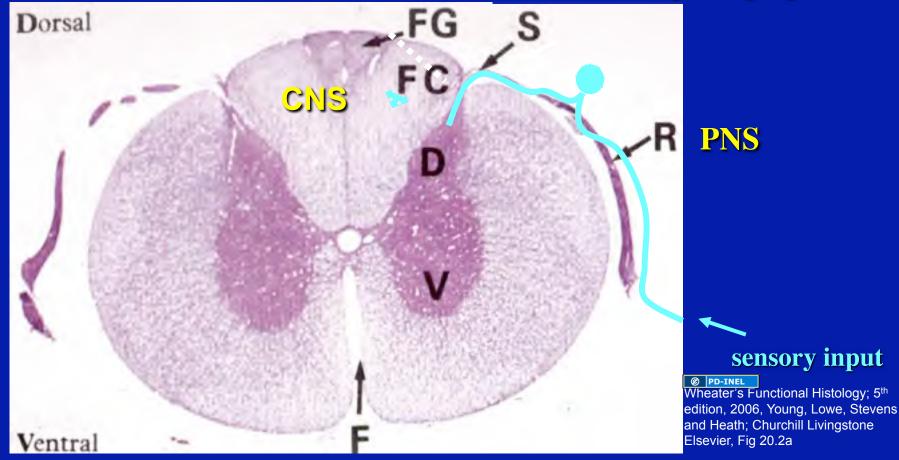


© PD-INEL From **Neuroscience** (2nd edition) by Dale Purves, et al. 2001 by Sinauer Associates, Inc <u>Figure 16.11</u>

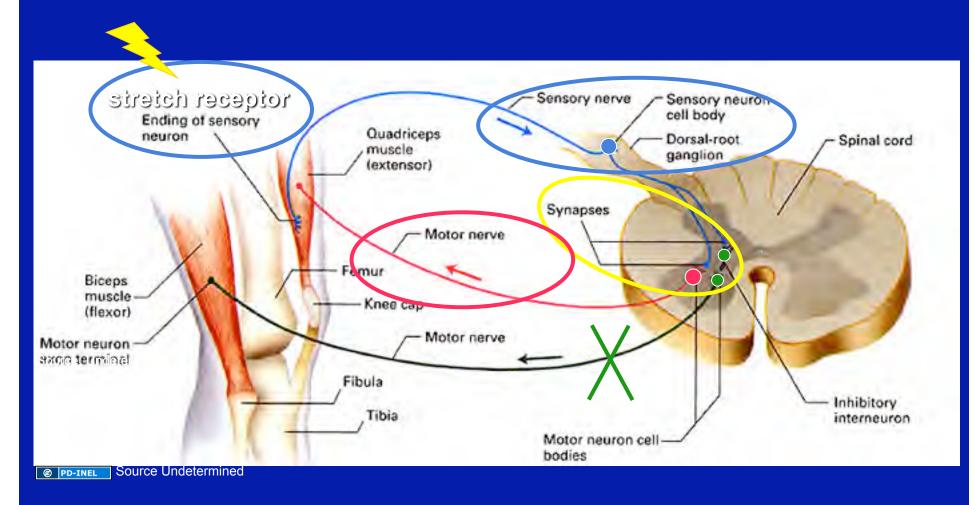


Somatic sensory neurons also have components in both CNS and PNS

pseudounipolar sensory neuron in a dorsal root (spinal) ganglion



Neuromuscular spindles are stretch receptors that regulate muscle tone via the spinal stretch reflex



Additional Source Information

for more information see: http://open.umich.edu/wiki/CitationPolicy

Slide 5: Basic Histology – Text & Atlas, 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-1 Slide 7: Basic Histology – Text & Atlas, 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-1 Slide 8: Basic Histology – Text & Atlas, 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-1

Slide 9: Julius Cornelius Schaarwächter, Wikipedia, http://commons.wikimedia.org/wiki/File:Wilhelm_von_Waldever-Hartz - 1891.jpg;

- Wheater's Functional Histology, 5th edition, 2006, Young, Lowe, Stevens and Heath, Churchill Livingstone Elsevier, Fig 7.4d Slide 10: Neuron to Brain, 3rd edition, 1992, Nicholls, Martin and Wallace, Sinauer, Fig 6
- Click 14. Henry Histolary, Ord center, 1992, Henry and Lyne Martin and Walacky, Chader,
- Slide 11: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6-1
- Slide 12: Color Atlas of Basic Histology, 1993, Berman, Appelton and Lange, Fig 6-4, University of Kansas Medical Center, Wikipedia, http://commons.wikimedia.org/wiki/Franz_Nissl
- Slide13: Cell and Tissue Ultrastructure A Functional Perspective, 1993, Cross and Mercer, Freeman and Co., Page 127
- Slide 14: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6-1
- Slide 15: Color Atlas of Histology, 1992, Erlandsen and Magney, Mosby Book, Fig 9-3
- Slide 16: Wheater's Functional Histology, 5th edition, 2006, Young, Lowe, Stevens and Heath, Churchill Livingstone Elsevier, Fig 7.5c
- Slide 17: Redrawn from Korean Neuroscience Newsletterhttp://aids.hallym.ac.kr/d/kns/tutor/medical/01premed2/chapter45/antret.html
- Slide 18: J.E. Lochner, M. Kingma, S. Kuhn, C.D. Meliza, B. Cutler, B.A. Scalettar, http://www.lclark.edu/~bethe/
- Slide 19: Originally from Duncan JE, Goldstein LSB (2006) The Genetics of Axonal Transport and Axonal Transport Disorders. PLoS Genet 2(9): Pages 1275ce -84., Reproduced from The Journal of Cell Biology, 2002, 157 (5) by copyright permission of The Rockefeller University Press; David Van Vactor, Havard University, http://focus.hms.harvard.edu/2004/sept3 2004/research briefs.html
- Slide 20: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig 6.7, Panel B courtesy of Olaf Mundigl and Pietro de Camilli in The Molecular Biology of the Cell by B. Alberts et al., 4th edition, 2002, Garland Science
- Slide 21: Modified from Health Education assets Library <u>http://www.healcentral.org/content/collections/McGill/7no11anim-450x580.swf</u>; Original Image: Astrid Vincent Andersen, <u>http://fantastrid.googlepages.com/anatomydrawings</u>; Cell and Tissue Ultrastructure – A Functional Perspective by Cross and Mercer, 1993, Freeman and Co. Page 135
- Slide 22: Wheater's Functional Histology, 5th edition, 2006, Young, Lowe, Stevens and Heath, Churchill Livingstone Elsevier, Fig 7.12a
- Slide 23: Color Atlas of Histology, 1992, Erlandsen and Magney, Mosby Book, Fig 8-18
- Slide 24: Photograph by AK Christensen from slide by Ray Truex, Dept of Anatomy, Temple Univ. School of Medicine
- Slide 25: Wikipedia, <u>http://en.wikivisual.com/index.php/;</u> Theodor_Schwann, Kelley, Kaye and Pawlina, "Histology, a Text and Atlas," 4th ed., page 284. Neuron-Ross4-284.tif.
- Slide 26: Basic Histology Text & Atlas; 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-30

Slide 27: Basic Histology – Text & Atlas, 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-30

Slide 28: Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 17.6 a; Source of Removed Image: Histology-A Text and Atlas by M.H. Ross and W. Pawlina, 5th edition, 2006, Lippincott Williams and Wilkins, Fig 12.11

Slide 29: Histology – A Text and Atlas, 5th edition, 2006, Ross and Pawlina, Lippincott Williams and Wilkins, Fig 12.11

Slide 30: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6.9, Tulane University Mata Medical Library

Slide 31: A.J. Lanterman: Ueber den feineren Bau der markhaltigen Nervenfasern. Arch. F. mikrosk. Anat. 1877 13:1-8; Modified from Histology – A Text and Atlas, 5th edition, 2006, Ross and Pawlina, Lippincott Williams and Wilkins , Fig 12.10b

Slide 32: Service De Neurologie, Hôpital Universitaire Dupuytren, Li, http://www.unilim.fr/neurolim/Images/NNFig08.jpg

- Slide 33: Wikimedia Commons, <u>http://commons.wikimedia.org/wiki/File:Louis-Antoine_Ranvier.jpg</u>; Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6.10
- Slide 34: Color Atlas of Histology, 1992, Erlandsen and Magney, Mosby Book, Fig 9-15
- Slide 35: Color Atlas of Histology, 1992, Erlandsen and Magney, Mosby Book, Fig 9.14
- Slide 36: Wheater's Functional Histology, 5th edition, 2006, Young, Lowe, Stevens and Heath, Churchill Livingstone Elsevier, Fig. 7.6 A, axon, S, Schwann cell nucleus.
- Slide 37: Japanese Kodachrome slide set, Slide 1084
- Slide 38: Color Atlas of Histology; 1992, Erlandsen and Magney, Mosby Book, Fig 9-13
- Slide 39: Source Undetermined
- Slide 40: Original Source Removed Modified from Neuroscience by D. Purves et al., 2001, 2nd ed., SinauerFig. 3.13, http://www.ncbi.nlm.nih.gov/books/bookres.fcgi/neurosci/ch3f13.gif
- Slide 41: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6-21
- Slide 42: Wheater's Functional Histology, 5th edition, 2006, Young, Lowe, Stevens and Heath, Churchill Livingstone Elsevier, Fig 7.5b
- Slide 43: Japanese slide set, Humio Mizoguti, Department of Anatomy, Kobe University School of Medicine, Slide #1091
- Slide 44: Color Textbook of Histology, 2nd edition, 1994, Gartner and Hiatt, Williams and Wilkins, Fig 7.5
- Slide 45: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6.20
- Slide 46: Basic Histology Text & Atlas, 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9.34
- Slide 47: Color Atlas of Basic Histology, 1993, Berman, Appelton and Lange, Fig 6.15
- Slide 48: Human Histology, 2nd edition, Stevens and Lowe, Mosby, Fig. 6.3
- Slide 49: Histology A Text and Atlas, 5th edition, 2006, Ross and Pawlina, Lippincott Williams and Wilkins , Plate 23 Slide 51: Source Undetermined
- Slide 52: Source Color Atlas of Basic Histology, 1993, Berman, Appelton and Lange, Fig 6-10
- Slide 53: Modified from Basic Histology Text & Atlas, 10th edition, 2003, Junqueira and Carneiro, Lange McGraw-Hill, Fig 9-38
- Slide 54: Japanese slide set, Humio Mizoguti, Department of Anatomy, Kobe University School of Medicine, Slides #1069a and #1069b
- Slide 55: Color Atlas of Histology, 1992, Erlandsen and Magney, Mosby Book, Fig 9-20

Slide 58: Netter's Essential Histology, 2008, Ovalle and Nahirney, Elsevier, Page 467

Slide 59: Wikimedia Commons, http://commons.wikimedia.org/wiki/File:C_Golgi.jpg; Wikipedia, http://en.wikipedia.org/wiki/Golgi_tendon_organ; From Neuroscience (2nd edition) by Dale Purves, et al. 2001 by Sinauer Associates, Inc Figure 16.11, http://www.ncbi.nlm.nih.gov/books/bv.fcgi?rid=neurosci.figgrp.1105

Slide 60: Color Textbook of Histology; 2nd edition, 1994; Gartner and Hiatt; Williams and Wilkins; Fig 71.

Slide 61: Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 20.2a Slide 62: Source Undetermined

Slide 56: Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.33a; Wikipedia, http://en.wikipedia.org/wiki/File:Wilhelm Kuhne.jpg

Slide 57: Wheater's Functional Histology; 5th edition, 2006, Young, Lowe, Stevens and Heath; Churchill Livingstone Elsevier, Fig 7.30b